

CASE REPORT

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A de novo whole gene deletion of XIAP detected by exome sequencing analysis in very early onset inflammatory bowel disease: a case report

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Abstract

Background: Children with very early-onset inflammatory bowel disease (VEO-IBD), those diagnosed at less than 5 years of age, are a unique population. A subset of these patients present with a distinct phenotype and more severe disease than older children and adults. Host genetics is thought to play a more prominent role in this young population, and monogenic defects in genes related to primary immunodeficiencies are responsible for the disease in a small subset of patients with VEO-IBD.

Case Presentation: We report a child who presented at 3 weeks of life with very early-onset inflammatory bowel disease (VEO-IBD). He had a complicated disease course and remained unresponsive to medical and surgical therapy. The refractory nature of his disease, together with his young age of presentation, prompted utilization of whole exome sequencing (WES) to detect an underlying monogenic primary immunodeficiency and potentially target therapy to the identified defect. Copy number variation analysis (CNV) was performed using the eXome-Hidden Markov Model. Whole exome sequencing revealed 1,380 nonsense and missense variants in the patient. Plausible candidate variants were not detected following analysis of filtered variants, therefore, we performed CNV analysis of the WES data, which led us to identify a *de novo* whole gene deletion in XIAP.

Conclusion: This is the first reported whole gene deletion in XIAP, the causal gene responsible for XLP2 (X-linked lymphoproliferative Disease 2). XLP2 is a syndrome resulting in VEO-IBD and can increase susceptibility to hemophagocytic lymphohistocytosis (HLH). This identification allowed the patient to be referred for bone marrow transplantation, potentially curative for his disease and critical to prevent the catastrophic sequela of HLH. This illustrates the unique etiology of VEO-IBD, and the subsequent effects on therapeutic options. This cohort requires careful and thorough evaluation for monogenic defects and primary immunodeficiencies.

Keywords: VEO-IBD (very early-onset IBD), XIAP (x-linked inhibitor of apoptosis), XLP2 (X-linked lymphoproliferative Disease 2), WES (whole exome sequencing)

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Background

Primary immunodeficiencies are a heterogeneous group of disorders that range in severity and clinical presentation, and may also lead to immune dysregulation such as severe inflammatory bowel disease (IBD), most frequently in patients with very early-onset IBD (VEO-IBD). VEO-IBD presents with a unique phenotype, including severe disease that is often unresponsive to conventional therapies. Disease causing variants have been detected in *IL-10*, *IL-10R* [1–4], *CYBB*, *CYBA*, *NCF1*, *NCF2* and *NCF4* (chronic granulomatous disease), *FOXP3*, *WAS*, *MEFV*, *ITGB2*, as well as other genes in patients with severe VEO-IBD [3, 5–7]. In addition, mutations have been identified in X-linked inhibitor of apoptosis (*XIAP*) in patients with VEO-IBD [8]. While defects in the gene *SH2D1A* were initially detected in patients with X-linked lymphoproliferative disease, known as XLP1 [9, 10], mutations in *XIAP* result in XLP2 [11]. IBD can be the first and only feature of XLP2. We report here a case of a male who presented in the neonatal period with severe refractory IBD, without any evidence of immunodeficiency, and was found to have a *de novo* whole gene deletion of *XIAP* at age 17. This is the first reported whole gene deletion identified in the literature and illustrates the unique pathogenesis of disease in children with VEO-IBD.

Case Presentation

The patient presented with VEO-IBD at 3 weeks of life with severe, bloody diarrhea. His initial endoscopy and colonoscopy, performed at 8 weeks, demonstrated upper tract disease (gastric and duodenal) and severe pancolitis. At 5 months of age, he developed a perianal sinus tract. He was initially treated with Azulfidine, steroids and elemental formula. The patient was unable to tolerate oral feeds and was thus TPN (total parental nutrition) dependent. He continued to have profound growth failure (<5 %) and due to lack of response to the addition of antibiotics and enemas to his regimen, a G-tube was placed and Azathioprine was initiated. His course remained severe, complicated by pathological vertebral fracture. Repeat endoscopy at 2 years of age demonstrated severe upper tract disease and worsening colonic disease. His perianal disease progressed as well, with abscess and fistula formation. At approximately 4 years of age he was found to have a distal rectal stricture requiring repeated dilations. He was initiated on infliximab at 5 years of age in addition to azathioprine as dual therapy, which was changed to methotrexate a year later. At 7 years of age, during a rectal dilation he developed a colonic perforation and required a diverting ileostomy. The patient continued to be medically refractory, therefore, when he was 10 years old he underwent a colectomy. Post-operatively, his disease remained severe, and was unresponsive to medical therapy, including

adalimumab, methotrexate, antibiotics, IVIG and finally vedolizumab. Repeat endoscopy performed demonstrated duodenal stricture and severe ileal disease (Fig. 1). In addition, the patient developed recurrent skin and intestinal abscesses and had significant steroid exposure throughout this time.

This patient's young age at diagnosis and progressive disease severity prompted an immunologic evaluation to detect primary immunodeficiencies associated with VEO-IBD. At age 16, further studies including quantitative immunoglobulins, vaccine titers, B and T cell lymphocyte subset analysis, natural killer (NK) cells, and toll like receptors were obtained. He was found to have normal levels of circulating T cells, but mildly low subsets of NK cells. His B cells exhibited activation as detected by CD80 expression and he had increased plasmablasts (4 %) and low IgM memory B cells (3 %).

In addition, when he was 17 years old, after written informed consent was obtained from his parents and assent from the patient, the patient and his family were recruited in the VEO-IBD research study using whole exome sequencing (WES) to detect rare or novel variants that contribute to disease at our institution.

Exome sequencing of the patient and his mother were performed under protocol 2002-07-2805 that was approved by the Institutional Review Board at The Children's Hospital of Philadelphia (CHOP). Exome capture was performed using the Agilent SureSelect V4 kit then sequenced using the Illumina HiSeq sequencer at an average coverage depth of 100X. Paired-end reads were aligned to the human reference genome GRCh37.p10 using Novoalign (V2.07.18; <http://www.novocraft.com>). Variants were detected using The Broad Institute's GATK [12] best practices, then annotated using SNPEff [13], 1000 Genomes Project (www.1000genomes.org/), Exome Variant Server (EVS) (<http://evs.gs.washington.edu/EVS/>), the Exome Aggregation Consortium (ExAC) (<http://exac.broadinstitute.org>; release 0.3), and the Combined Annotation-Dependent Depletion tool (CADD) [14]. Identified variants were then filtered, retaining only nonsynonymous missense and nonsense, rare (minor allele frequency <0.1 % in all 3 databases) or novel mutations, with a CADD score ≥ 10 . Variants were further restricted to genes involved in primary immunodeficiency. Copy number variation (CNV) analysis was performed using the eXome-Hidden Markov Model (XHMM) [15].

Genotyping was conducted using the Illumina HumanHap550v3.0 Beadchip. The data was analyzed using Beadstudio software (Illumina) to confirm the deletion that was found in the WES data.

The *XIAP* CNV was confirmed using the Bio-Rad QX100 droplet digital PCR (ddPCR) platform. Pre-designed primer/probe configurations were used for *XIAP*

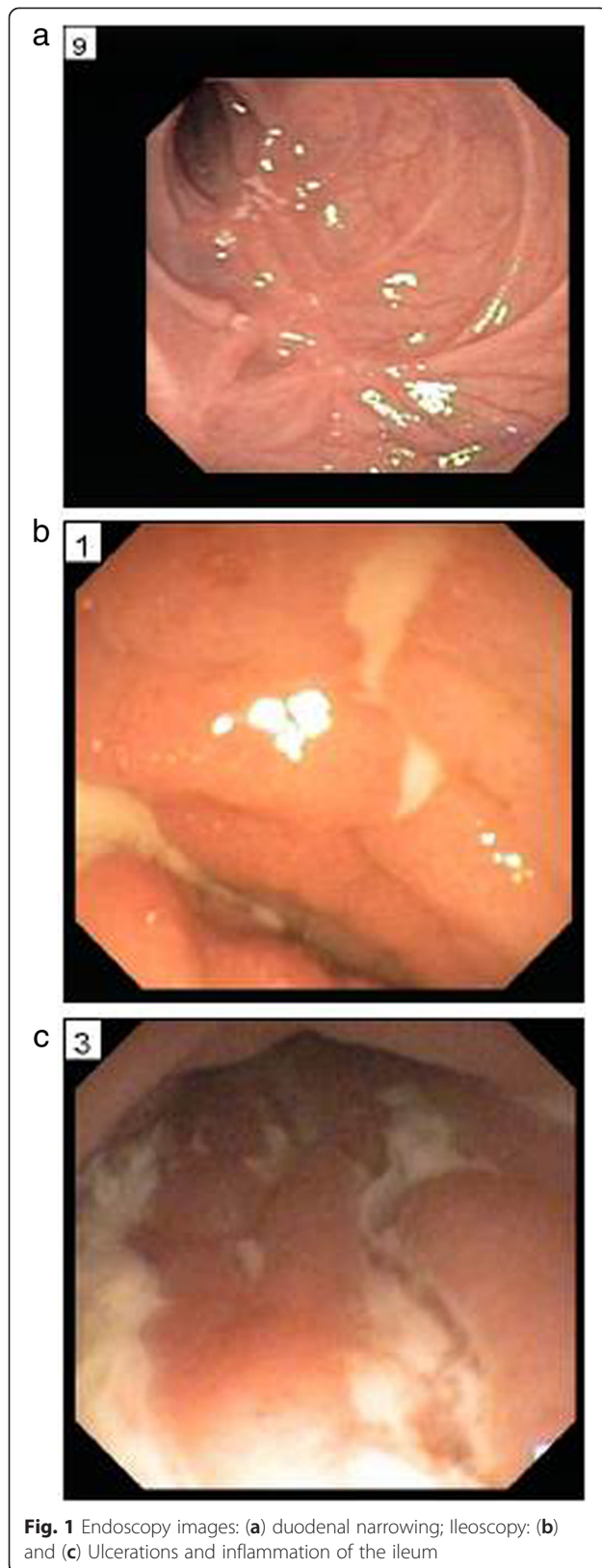


Fig. 1 Endoscopy images: (a) duodenal narrowing; ileoscopy: (b) and (c) Ulcerations and inflammation of the ileum

(cat#: Hs04517714_cn) and the reference gene *TERT* (cat#4403326). Standard ddPCR conditions and analysis methods were used for this assay [16].

Whole exome sequencing revealed 1,380 nonsense and missense variants in the patient. However, after restricting the variants to rare or novel variants in genes associated with immunodeficiency, only a single variant was identified in *CCHCR1* (p.Gln609Pro) that met our criteria. The same variant was also found in the patient's unaffected mother and therefore was unlikely to be associated with his disease.

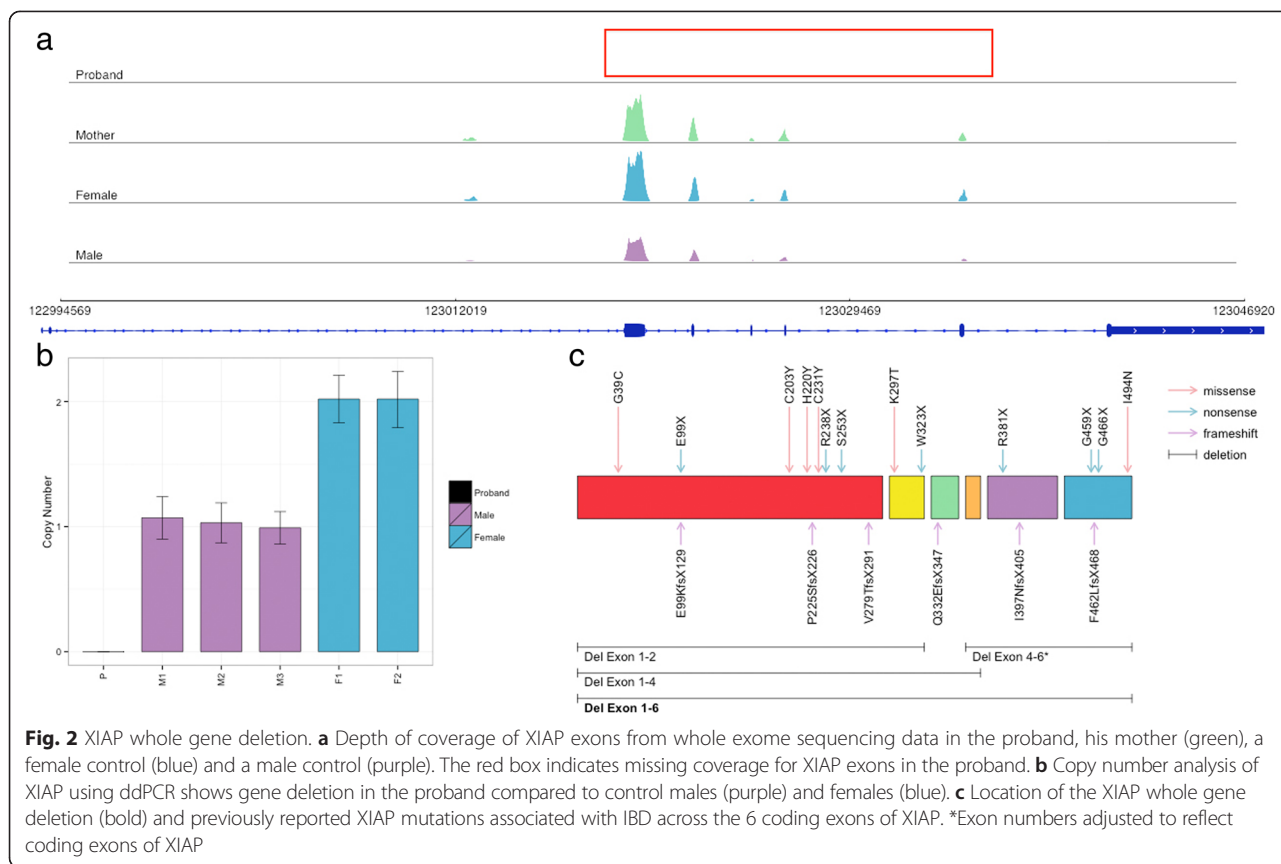
We then performed CNV analysis of the WES data of the patient and his mother, which led us to identify a *de novo* whole gene deletion in *XIAP* (Fig. 2a). The deletion was then validated using both SNP arrays and ddPCR (Fig. 2b). The SNP array deletion spanned from rs5958318 at position chrX:122,992,832, near the 5' end to rs10521711 at position chrX:123,047,926 near the 3' UTR of *XIAP*, which confirmed that the entire gene was deleted.

Functional analysis of *XIAP* demonstrated lack of *XIAP* expression in CD4+ and CD8+ T cells, NK cells and B cells. Following identification of this deletion the patient was referred to our Bone Marrow Transplant Team for a bone marrow transplantation (BMT).

Discussion

Monogenic defects of primary immunodeficiency can present as severe neonatal IBD, including epithelial barrier defects, phagocyte defects (Chronic granulomatous disease, defects of NADPH oxidase LAD), B and T cell abnormalities (Severe combined immunodeficiency, common variable immunodeficiency, Wiskott-Aldrich Syndrome, immunodysregulation polyendocrinopathy enteropathy X-linked syndrome), autoimmunity (including XLP2) and IL-10R defects. Due to this patient's severe refractory disease and age of onset, evaluation for primary immunodeficiency was performed. Although the immune work up was unrevealing, evaluation of WES data demonstrated a whole gene deletion of *XIAP*.

X-linked lymphoproliferative disease, XLP1, first described in 1974 by Purtilo, results in the immunodeficiency of severe susceptibility to EBV due to defects in NK cell mediated toxicity, and leads to hemophagocytic lymphohistocytosis (HLH), dysgammaglobulinemia and lymphoma [17]. This triad is secondary to inactivating mutations in *SH2D1A*, encoding SLAM-associated protein (SAP), on chromosome Xq25 [10]. In 2006, Rigaud et al. demonstrated that mutations in the gene that encodes the X-linked inhibitor-of-apoptosis *XIAP* (also termed *BIRC4*) resulted in another X-linked lymphoproliferative syndrome, XLP2, distinct from SAP deficiency XLP (XLP1) [11]. The immune dysregulation in these patients, including our patient described here, was characterized by absent *XIAP* in lymphocytes, NK cells and



myeloid cells. Two of the patients described by Riguard had hemizygous mutations in *XIAP* and developed IBD. Worthey et al. first identified *XIAP* deficiency in VEO-IBD, in a 15-month-old male with severe fistulizing Crohn’s disease. This patient had a novel hemizygous mutation and ultimately underwent BMT, which proved to be curative [8]. Since then, several other cases of *XIAP* missense or nonsense mutations in patients with VEO-IBD have been reported [18]. Zeissig et al. further demonstrated the role of *XIAP* deficiency in severe VEO-IBD, with the identification of variants in 4 male pediatric patients, ages 1–16 [18]. The variant identified in the patient described in this study is the first whole gene deletion, to our knowledge, to be reported in VEO-IBD, and its identification provided a potentially curative therapeutic option. The mapping of the deletion described in this study compared to mutations previously reported in the literature in association with IBD is depicted in Fig. 2c.

The inhibitors of apoptosis (IAPs) genes, such as *XIAP*, are instrumental for both innate and adaptive immunity. *XIAP* inhibits apoptosis in activated T cells, thus allowing expansion and survival via blockade of initiator and effector caspases [19] or through binding to death-inducing caspases and subsequent proteosomal mediated degradation. IAPs contain 1–3 baculoviral IAP repeat (BIR)

homology domains, a ubiquitin binding domain (UBA) and a C-terminal RING domain [20]. BIR2 and BIR3 domains of cIAP1, CIAP2 and *XIAP* are required for binding to and suppression of specific cell death-inducing caspases [19]. *XIAP* inhibits apoptosis by binding to and blocking activated forms of caspases 3,7 and 9. *XIAP* participates in innate immunity through signaling pathways and cellular responses via ubiquitin ligases activity [20]. It is required in the signaling and function of the pattern recognition receptors, *NOD1*, *NOD2* and Dectin-1 [19]. Upon activation during infection, *NOD1/2* and Dectin-1 promote cytokine production, with subsequent clearance of pathogens. *XIAP* is also involved in the inhibitory signaling of inflammasomes (*NLRP3* in murine models) and decreases TNF- α activation by pathogen-associated molecular patterns (PAMPs). Therefore, *XIAP* eradicates local acute inflammation and maintains low levels of baseline inflammation. In *XIAP*-deficient patients, both arms of the immune system, innate and adaptive responses, are affected, with a cyclical environment of unchecked activation of inflammasomes and intestinal permeability to pathogens leading to the accumulation of pro-inflammatory cytokines such as TNF- α , IL-1 β and IL-18. Finally, there is increased cell-death of cells, including lymphocytes and myeloid cells and chronic inflammation. Ultimately the patient develops the phenotype of splenomegaly, IBD and HLH [20].

Conclusion

Due to the extensive differential in the work up of our patient with severe VEO-IBD, next generation sequencing technology can be of benefit. Targeted sequencing, to evaluate for identified monogenic defects in VEO-IBD, or WES to study the whole exome, can offer a quicker method of analysis rather than a gene-by-gene approach. The results of this critical finding directed the course of action of treatment. Because XIAP deficiency has a high risk of fatal HLH, and due to his severe disease, he has undergone bone marrow transplantation. While XLP2 functional analysis currently exists, the wide differential in this case made WES, and the multidisciplinary approach involving gastroenterology, genetics, immunology and bioinformatics an attractive and effective option.

Consent

Written informed consent was obtained from the patient's parents as he is a minor, for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

Abbreviations

BMT: bone marrow transplantation; CADD: combined annotation-dependent depletion; CHOP: The Children's Hospital of Philadelphia; CNV: Copy number variation analysis; DHR: dihydrochlorodamine; EVS: Exome Variant Server; ExAC: Exome Aggregation Consortium; HLH: hemophagocytic lymphohistiocytosis; IAP: inhibitors of apoptosis; IBD: inflammatory bowel disease; MAF: minor allele frequency; NK: natural killer cells; PAMP: pathogen-associated molecular patterns; TPN: total parental nutrition; Sap: SLAM-associated protein; VEO-IBD: very early-onset inflammatory bowel disease; WES: whole exome sequencing; XHMM: eXome-Hidden Markov Model; XIAP: X-linked inhibitor-of-apoptosis; XLP1: X-linked lymphoproliferative disease; XLP2: X-linked lymphoproliferative Disease 2.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors contributed to this study, in particular. Study concept and design: Kelsen, Dawany, Piccoli, Mamula, Baldassano, Sullivan, Devoto. Acquisition of data: Kelsen, Martinez, Rappaport, Maurer, Grochowski. Analysis and interpretation of data: Kelsen, Dawany, Rappaport, Sullivan, Baldassano, Devoto, Maurer, Grochowski. Drafting of the manuscript: Kelsen, Dawany, Baldassano, Mamula, Sullivan, Devoto. Critical Revision of Analysis of the manuscript for important intellectual content: Kelsen, Dawany, Piccoli, Mamula, Sullivan, Baldassano, Devoto. Statistical analysis: Kelsen, Dawany, Devoto. Technical or material support: Dawany, Rappaport. Study Supervision: Kelsen, Baldassano, Devoto. All authors read and approved the final manuscript.

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Disclosures

None of the authors have disclosures related to this manuscript.

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References

- Glocker EO, Kotlarz D, Boztug K, et al. Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. *N Engl J Med*. 2009;361:2033–45.
- Glocker EO, Frede N, Perro M, et al. Infant colitis—it's in the genes. *Lancet*. 2010;376:1272.
- Glocker E, Grimbacher B. Inflammatory bowel disease: is it a primary immunodeficiency? *Cell Mol Life Sci*. 2012;69:41–8.
- Moran CJ, Walters TD, Guo CH, et al. IL-10R polymorphisms are associated with very-early-onset ulcerative colitis. *Inflamm Bowel Dis*. 2013;19:115–23.
- Dupuis-Girod S, Medioni J, Haddad E, et al. Autoimmunity in Wiskott-Aldrich syndrome: risk factors, clinical features, and outcome in a single-center cohort of 55 patients. *Pediatrics*. 2003;111:e622–7.
- Dhillon SS, Fattouh R, Elkadri A, et al. Variants in nicotinamide adenine dinucleotide phosphate oxidase complex components determine susceptibility to very early onset inflammatory bowel disease. *Gastroenterology*. 2014;147:680–9. e2.
- Muise AM, Xu W, Guo CH, et al. NADPH oxidase complex and IBD candidate gene studies: identification of a rare variant in NCF2 that results in reduced binding to RAC2. *Gut*. 2012;61:1028–35.
- Wortheley EA, Mayer AN, Syverson GD, et al. Making a definitive diagnosis: successful clinical application of whole exome sequencing in a child with intractable inflammatory bowel disease. *Genet Med*. 2011;13:255–62.
- Coffey AJ, Brooksbank RA, Brandau O, et al. Host response to EBV infection in X-linked lymphoproliferative disease results from mutations in an SH2-domain encoding gene. *Nat Genet*. 1998;20:129–35.
- Sayos J, Wu C, Morra M, et al. The X-linked lymphoproliferative-disease gene product SAP regulates signals induced through the co-receptor SLAM. *Nature*. 1998;395:462–9.
- Rigaud S, Fondaneche MC, Lambert N, et al. XIAP deficiency in humans causes an X-linked lymphoproliferative syndrome. *Nature*. 2006;444:110–4.
- McKenna A, Hanna M, Banks E, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res*. 2010;20:1297–303.
- Cingolani P, Platts A, Le Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly (Austin)*. 2012;6:80–92.
- Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM, Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet*. 2014;46:310–5.
- Fromer M, Purcell SM. Using XHMM software to detect copy number variation in whole-exome sequencing data. *Curr Protoc Hum Genet*. 2014; 81:7 23 1–7 1.
- Tsai EA, Grochowski CM, Falsey AM, et al. Heterozygous deletion of FOXA2 segregates with disease in a family with heterotaxy, panhypopituitarism, and biliary atresia. *Hum Mutat*. 2015;36:631–7.
- Purtilo DT, Cassel C, Yang JP. Letter: Fatal infectious mononucleosis in familial lymphohistiocytosis. *N Engl J Med*. 1974;291:736.
- Zeissig Y, Petersen BS, Milutinovic S, et al. XIAP variants in male Crohn's disease. *Gut*. 2015;64:66–76.
- Pedersen J, LaCasse EC, Seidelin JB, Coskun M, Nielsen OH. Inhibitors of apoptosis (IAPs) regulate intestinal immunity and inflammatory bowel disease (IBD) inflammation. *Trends Mol Med*. 2014;20:652–65.
- Aguilar C, Latour S. X-linked inhibitor of apoptosis protein deficiency: more than an X-linked lymphoproliferative syndrome. *J Clin Immunol*. 2015;35(4):331–38.